

# LYOPHILISED TETRA $^{TM}$ 2X qPCR MASTERMIX

# qPCR HANDBOOK

For Research Use Only. Not intended for diagnostic use.

CAT NO.: YSL-qP-TMM-100/500 100 reactions / 500 reactions

VERSION 5.0





### INTENDED USE

YouSeq Tetra™ MasterMix is a 2X ready-to-use mix enabling superior assay performance across various sample types. It is intended for use by trained professionals in a suitable molecular biology laboratory.

### CONTENTS

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Component	Cap Colour	Pack Size	
Component	Cap Colour	100 reactions	500 reactions
Lyophilised Tetra™ 2X qPCR MasterMix		1 x 1.1 mL*	5 x 1.1 mL*
MasterMix Resuspension Buffer (MMRB)		1 x 1.5 mL	5 x 1.5 mL
ROX Passive Reference Dye		1 x 10 μL	3 x 10 μL
DNase/RNase Free Water		1 x 1.5 mL	1 x 1.5 mL

<sup>\*</sup> Supplied lyophilised - requires resuspension. See instructions in resuspension section.

# RECOMMENDED ADDITIONAL REAGENTS & MATERIALS

qPCR primer/probe mixes (Recommended for use with YouSeq extensive qPCR design catalogue: www.youseq.com).

Nucleic acid template.

General laboratory equipment (pipettes, pipette tips, (micro)centrifuge tubes, etc.)

qPCR instrument.

### BEST PRACTICE

### Decontamination:

Before beginning laboratory work, thoroughly decontaminate any work surfaces & pipettes being used, to eliminate potential contamination.

### General use & set-up:

All components should be fully defrosted with contents at the bottom of the tube before opening. To ensure contents are at the bottom, centrifuge or gently tap the tube. After use, reagents should be returned to the freezer.

After resuspending or combining reagents, the tube should be pulse vortexed to ensure it is mixed well.

It is advised to set up the plate and reaction mix on ice to minimise artefact formation, which may reduce sensitivity.

When preparing the qPCR reaction mix, it is recommended to incorporate an overage when calculating the total number of reactions to compensate for potential volume losses incurred during pipetting.

### Set-up environments:

It is best practice to set up qPCR tubes/plates in two different environments - a clean (no template) lab and PCR (template) lab.



# BENCH SIDE PROTOCOL

### RESUSPENSION

Before first use, resuspend the designated component with the correct reagent and specified volume, as per the table below:

1. Add the resuspension reagent and pulse vortex the vial to ensure each is mixed well.

Component	Reagent	Volume	Location
Lyophilised Tetra™ 2X qPCR MasterMix	MMRB	1.1 mL	Clean lab

# ROX (INSTRUMENT DEPENDENT)

ROX can be required for instruments that use ROX as a passive reference, however, if using a qPCR primer/probe mix labelled with a ROX fluorophore, the passive reference dye should not be used. The table below outlines the qPCR instruments that require the addition of ROX to the MasterMix.

- 1. If ROX is required:
  - a. Dilute the ROX supplied according to the table below based on the intended qPCR instrument to be used.
  - b. Directly transfer 5  $\mu L$  of prepared ROX to the fully resuspended Tetra MasterMix.

Level of ROX	Instruments	Step 1: Volume of water to add to each ROX tube	Step 2: Add to MasterMix vial
High ROX Instruments	Applied Biosystems 7700, 7000, 7900, 7300, StepOne, StepOne Plus	No Dilution Required	5 μL
Low ROX Instruments	Applied Biosystems 7500 & 7500 FAST, ViiA7, Quantstudio, Stratagene MX	130 μL	5 μL
ROX Not Required:	All Other Instruments	Not Required	Not Required

# qPCR REACTION SET-UP

 $\mathsf{Tetra}^\mathsf{TM}$  qPCR MasterMix is supplied at 2X concentration. When setting up the qPCR reaction mix, the MasterMix is required to be half the total reaction volume.

Example: 10 μL of Tetra<sup>TM</sup> 2X qPCR MasterMix in a 20 μL total reaction volume.



# qPCR AMPLIFICATION PROTOCOL

1. Load the plate onto the qPCR machine and set-up the recommended qPCR protocol below.

Please note: If using a qPCR machine that uses ROX as a passive reference and a primer/probe mix containing a ROX labelled probe, ensure the passive reference is turned off or set to "none" before starting the run.

Step	Temperature	Time	Number of Cycles	
Initial Denaturation	95°C	3 minutes	-	
Denaturation	95°C	15 seconds	- x 45	
Annealing/Extension	60°C <b></b>	60 seconds		

- Collect fluorogenic data through the required channels during this step.
- 2. Start the run.

The recommended qPCR protocol above can be adjusted following the details below to optimise the reaction for the intended application.

### Uracil-DNA Glycosylase (UNG/UDG)

The Tetra<sup>TM</sup> 2X qPCR MasterMix contains dUTP's and Uracil-DNA Glycosylase (UNG/UDG). The enzyme will degrade DNA templates containing Uracil bases, in order to prevent carry-over contamination during PCR setup. However, it is designed to denature completely at 55°C and hence will not impact the PCR amplification.

#### Denaturation

Both denaturation steps at 95°C are recommended. However, when using a template with high GC content, this may be increased to 98-100°C to improve results.

### Annealing/Extension

The recommended annealing temperature is 60°C. The annealing temperature may be optimised by performing a temperature gradient within the annealing step between 60°C and 70°C to remove non-specific product.



# PRODUCT SPECIFICATIONS

### Storage

Store at -20°C from arrival. The products shelf life is outlined as an expiry date on the pouch label.

To reduce excessive freeze-thaws (<10), the MasterMix can be stored in small aliquots if required.

### Regulatory status

This product has been developed for Research Use Only and is not intended for diagnostic use.

### **Quality Control**

In accordance with the YouSeq Ltd ISO EN 13485-certified Quality Management System, each lot of YouSeq Lyophilised Tetra™ 2X qPCR MasterMix is tested against predetermined specifications to ensure consistent product quality.

#### **Technical Assistance**

For customer support, please contact:

e-mail: support@youseq.com phone: +44 (0)333 577 6697

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